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Genomic footprints of activated telomere maintenance mechanisms in cancer

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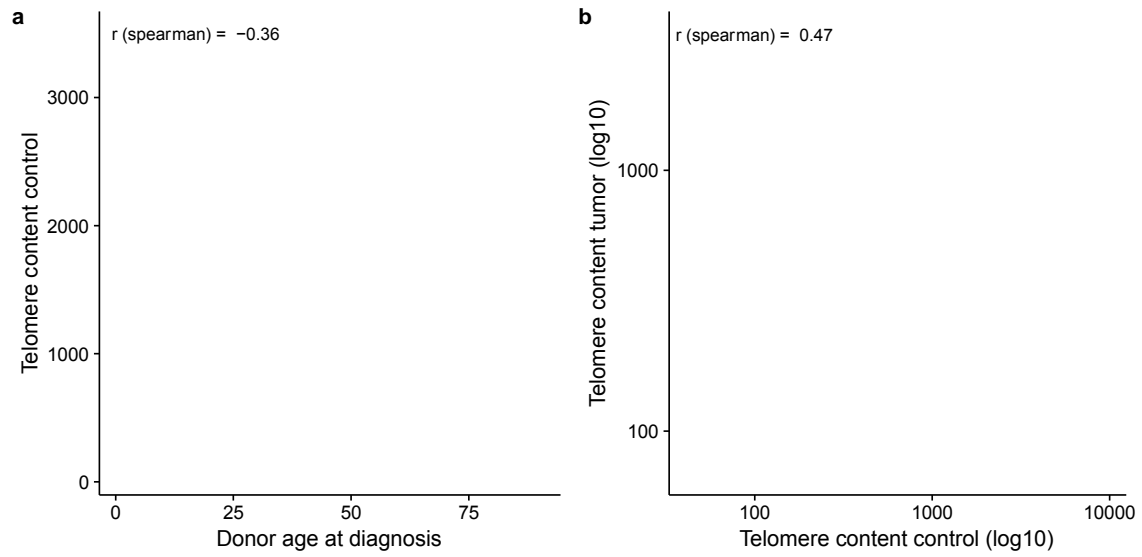
Supplementary Information

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Sieverling *et al.*

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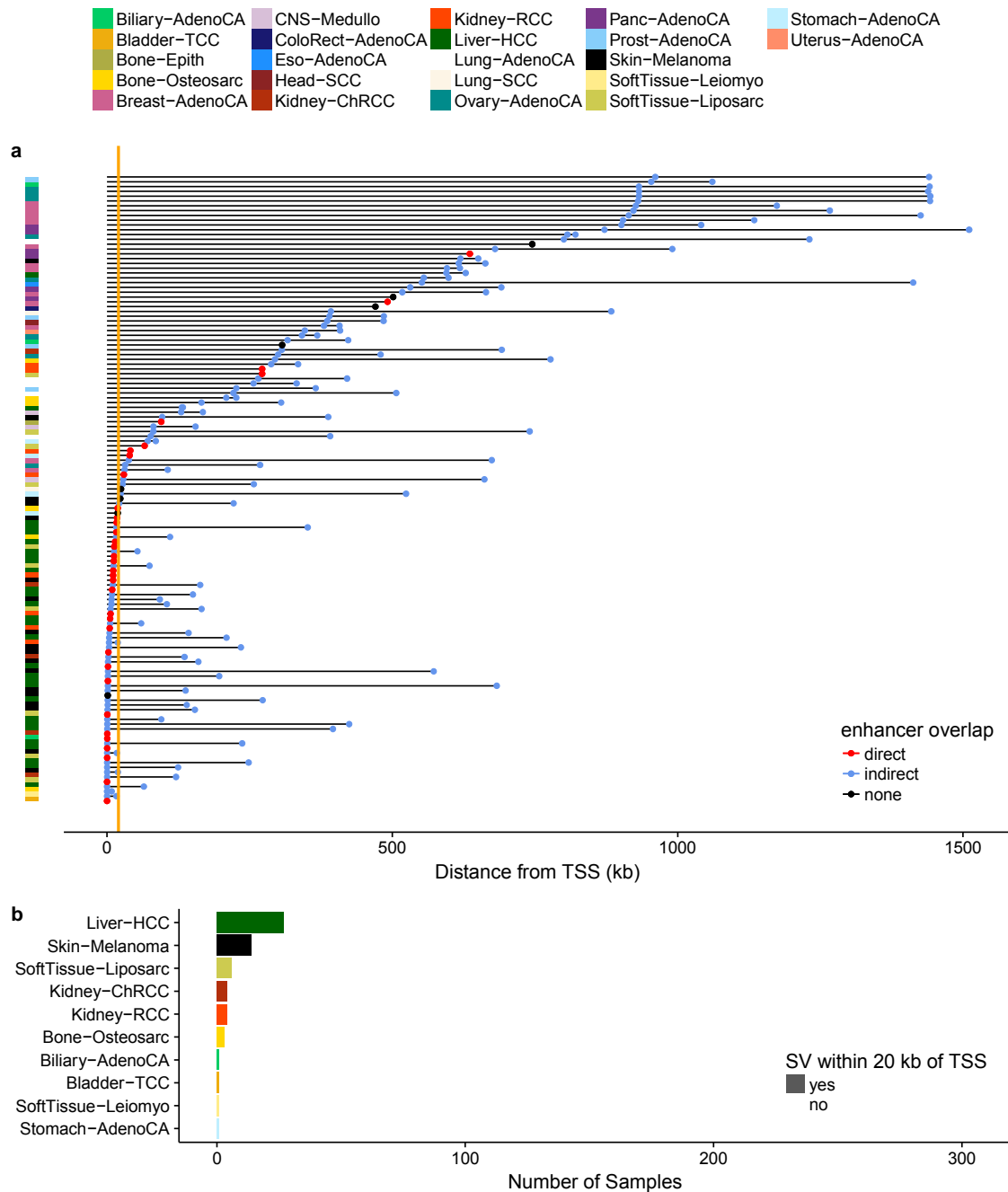
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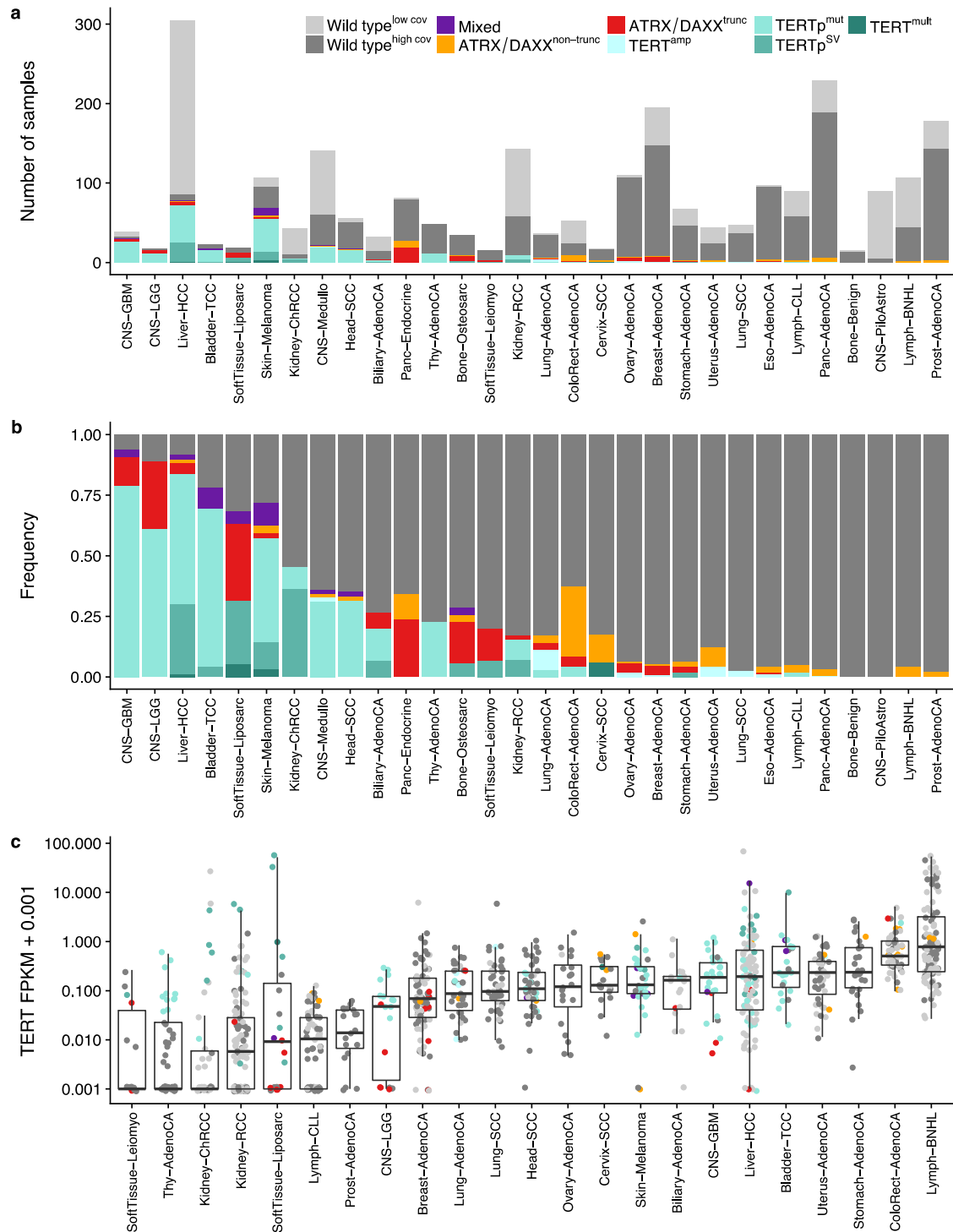
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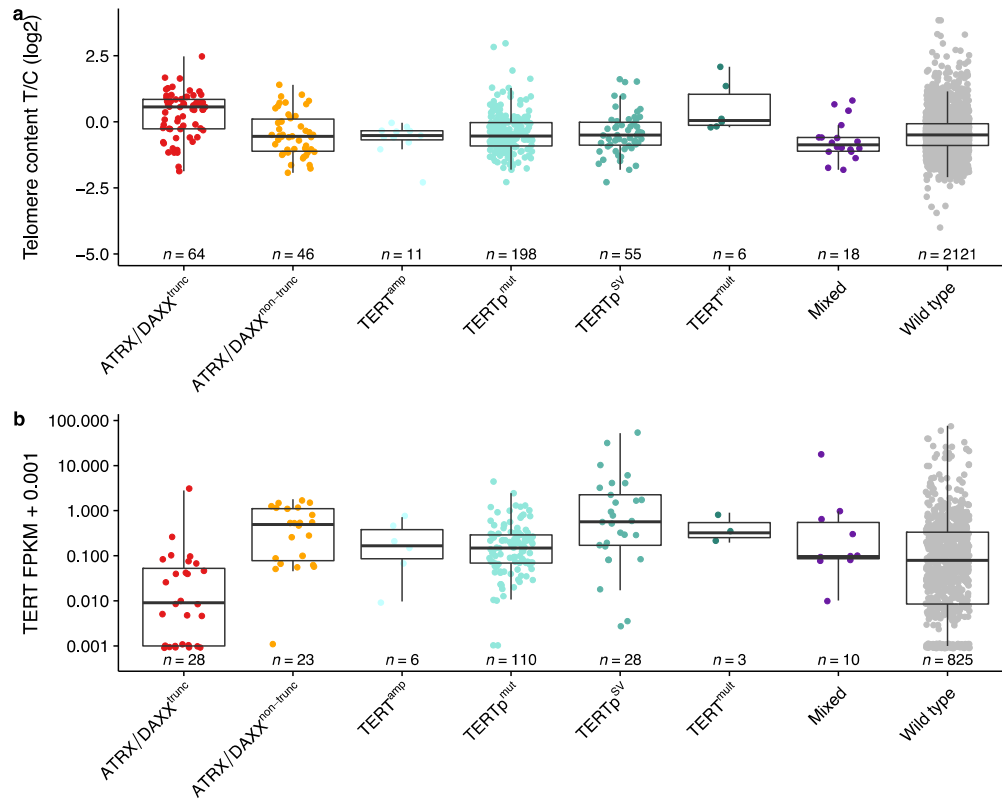
Supplementary Figure 1: Influences on telomere content. (a) Correlation of control telomere content and the patient age at diagnosis. (b) Correlation of telomere content in the tumor and the control sample.



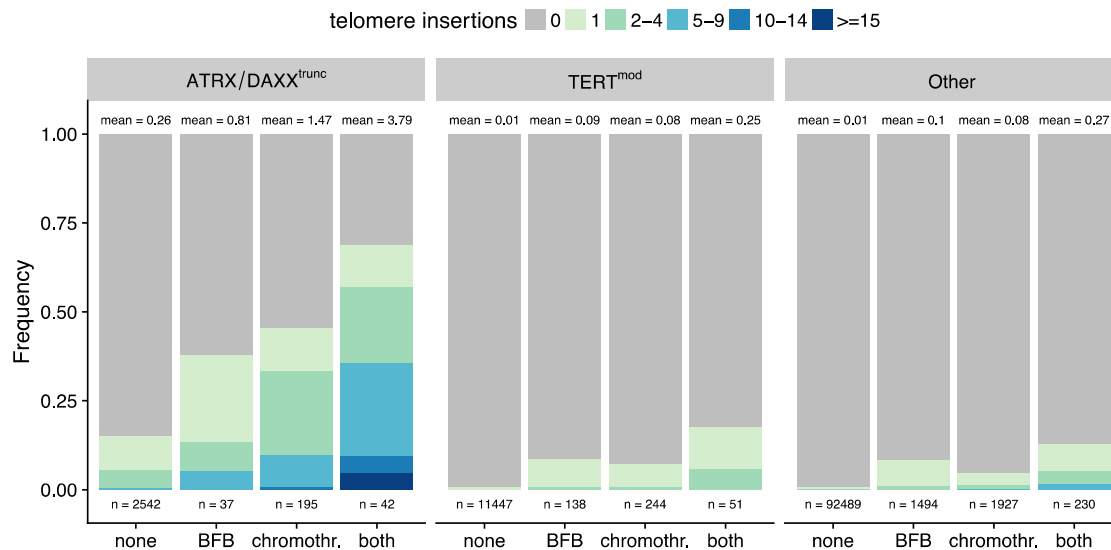
Supplementary Figure 2: Structural variations upstream of *TERT*. (a) Distance of structural variations (SVs) up to 1 mb upstream of the *TERT* transcription start site (TSS). For each tumor sample, only the SV closest to the TSS is shown. Direct overlaps of juxtaposed positions with dbSUPER enhancer regions are indicated in red. dbSUPER enhancers upstream of the SV are shown in blue, where the first point of each line is the position of the SV and the second point is the rearranged enhancer position. All tumor samples with SVs within 20 kb of the *TERT* TSS (orange line) were considered as *TERT*^{mod} for the further analysis. (b) Number of samples per tumor type with and without an SV within 20 kb of the *TERT* TSS. Only tumor types with at least one affected sample are shown.



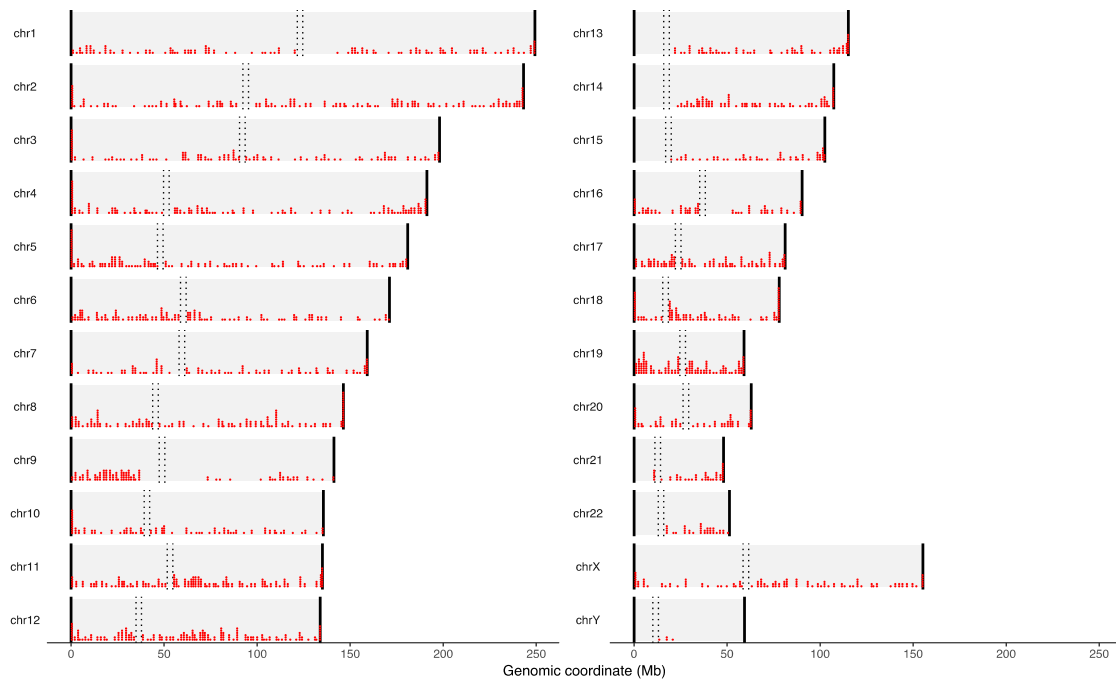
Supplementary Figure 3: TMM-associated mutations and *TERT* expression in different tumor types. (a) Number of samples with TMM-associated mutations. Wild type tumors (i.e. tumors without *TERT*^{mod} or mutation in *ATRX* or *DAXX*) were divided into those with a coverage of at least 10 reads at *TERT* promoter positions chr5:1,295,228 and chr5:1,295,250 (Wild type^{high cov}) and those without (Wild type^{low cov}). The tumor types are sorted by the frequency of ATRX/DAXX^{trunc} and TERT^{mod} samples. (b) Frequency of TMM-associated mutations. Wild type tumors with low coverage at the *TERT* promoter were excluded. (c) *TERT* expression in different tumor types. The tumor types are sorted by increasing median *TERT* expression. The center line of the boxplot is the median, the bounds of the box represent the first and third quartiles, the upper and lower whiskers extend from the hinge to the largest or smallest value, respectively, no further than 1.5 * IQR from the hinge (where IQR is the inter-quartile range, or distance between the first and third quartiles).



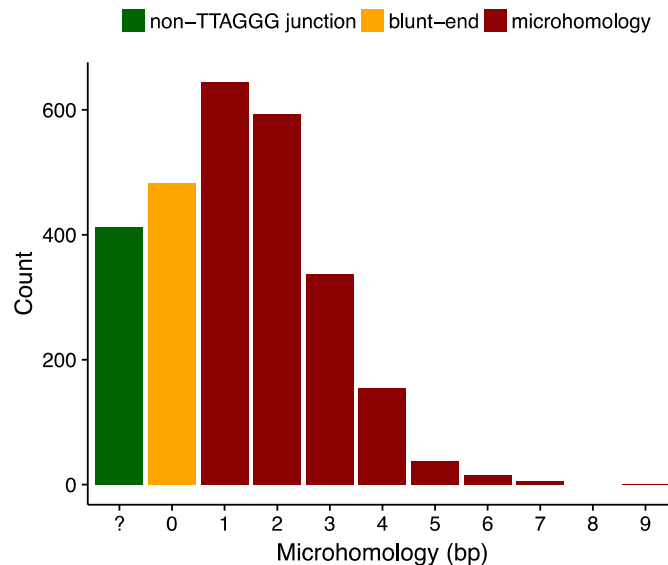
Supplementary Figure 4: Telomere content and *TERT* expression of tumor samples with different TMM-associated mutations. (a) Telomere content tumor/control log2 ratios. (b) *TERT* expression in FPKMs. The center line of the boxplot is the median, the bounds of the box represent the first and third quartiles, the upper and lower whiskers extend from the hinge to the largest or smallest value, respectively, no further than 1.5 * IQR from the hinge (where IQR is the inter-quartile range, or distance between the first and third quartiles).



Supplementary Figure 5: Co-occurrence of telomere insertions, breakage-fusion-bridge (BFB) cycles and chromothripsis on the same chromosome arm. Autosomal chromosome arms of patients with different TMM-associated mutations were divided into those with indication of BFB cycles, chromothripsis, both or none of these complex structural abnormalities. The frequency of telomere insertion counts is shown for each of the classes. The total number of chromosome arms falling into each category (n) and the mean number of telomere insertions is indicated.

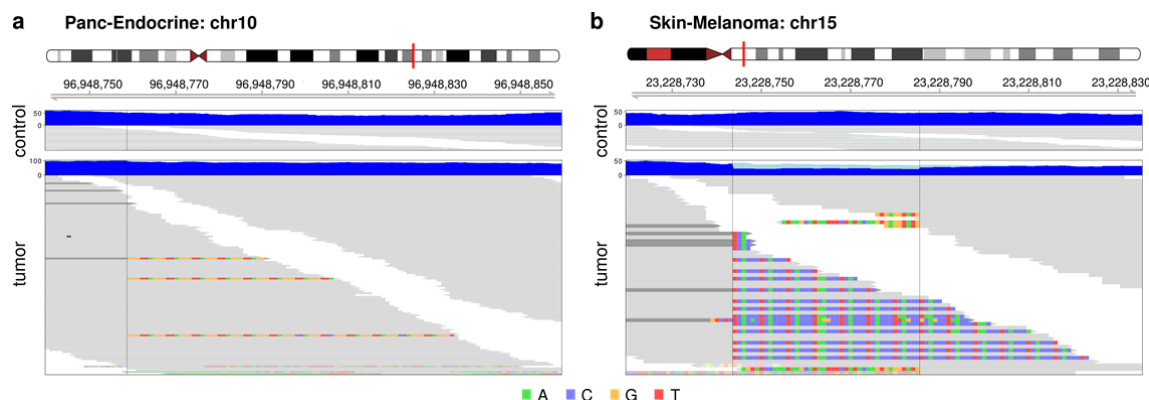


Supplementary Figure 6: Chromosomal positions of telomere insertions. Telomere insertions of all patients are shown.



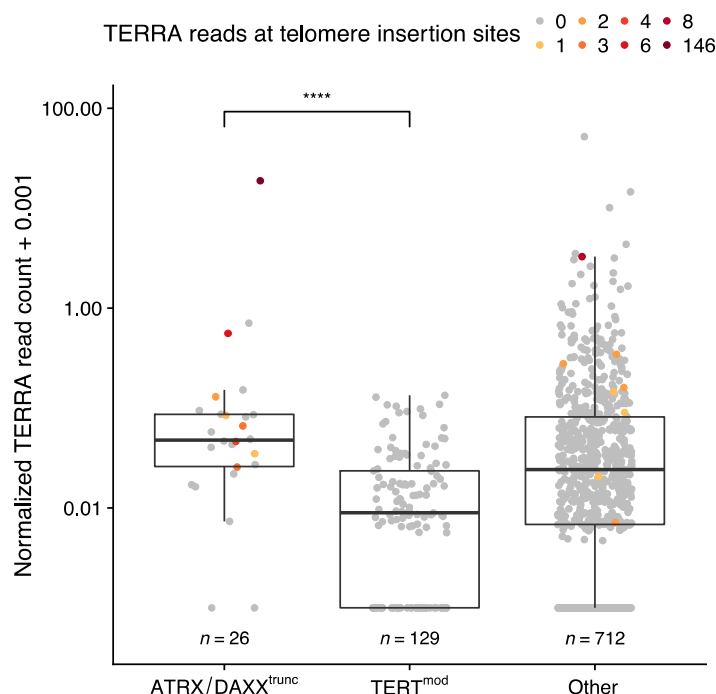
Supplementary Figure 7: Patterns of microhomology at telomere insertions. The number of homologous bases between the canonical TTAGGG telomere repeat and the human reference genome at telomere insertions is shown on the x-axis. The number of telomere insertions with a pattern of TTAGGG microhomology (red), blunt-end DNA joining (yellow) or without TTAGGG repeats at the junction site (green) are shown on the y-axis.

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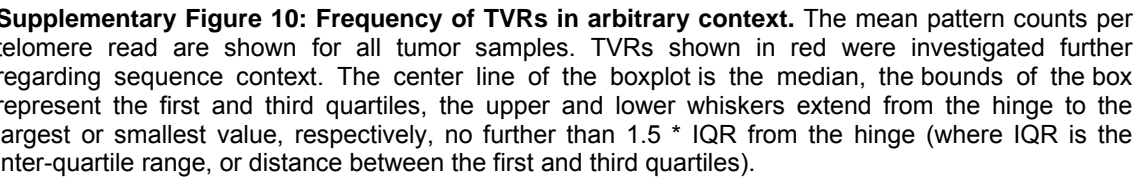
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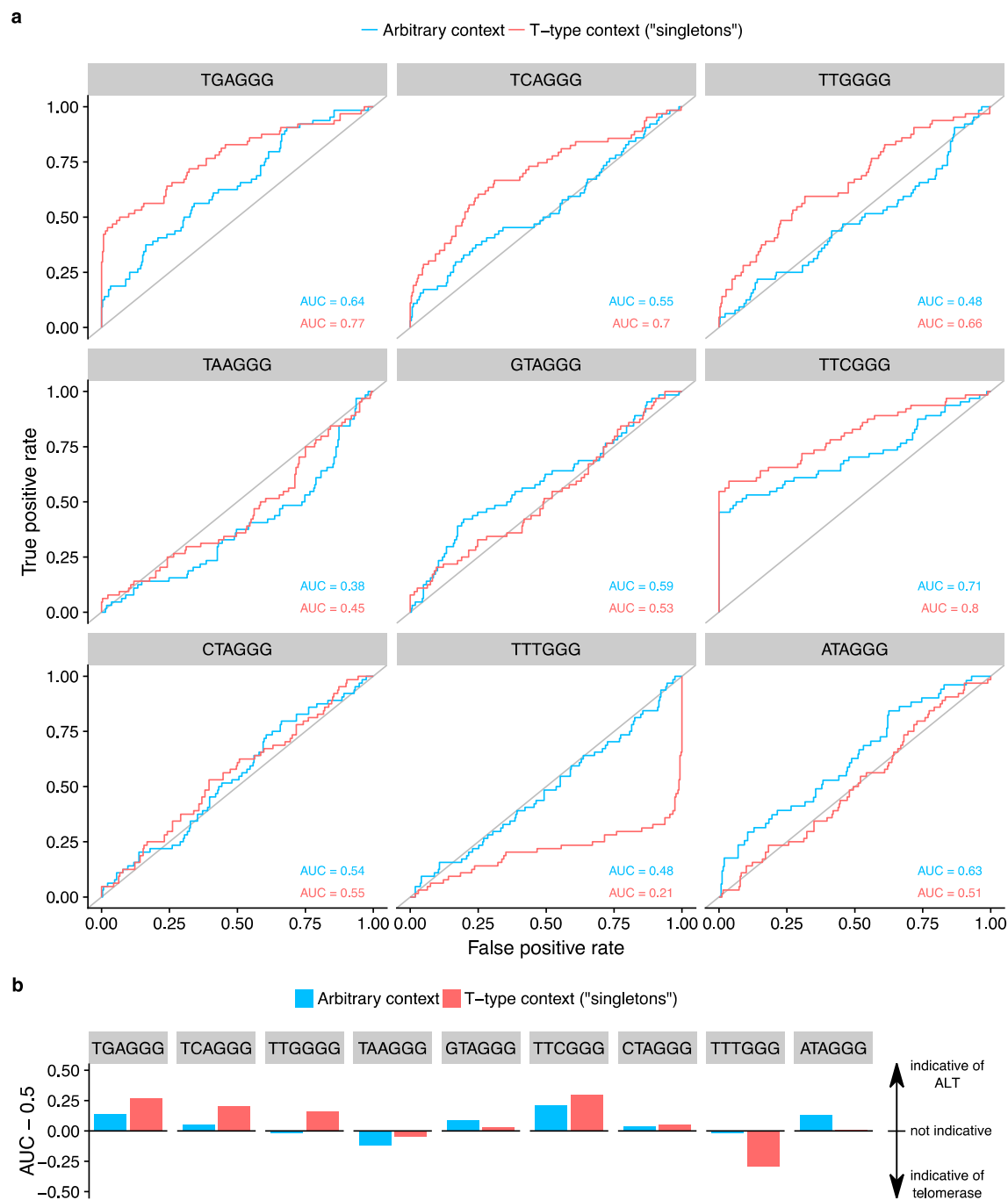
Supplementary Figure 8: Examples of one-sided telomere insertions without annotated accompanying structural variations. (a) Subclonal telomere insertion in pancreatic endocrine tumor sample SP102547 (copy number at position = 2; tumor purity = 0.87, as determined by copy number calls). Blue tracks show the sequencing coverage; light blue represents clipped sequences. Individual reads are grey and clipped bases are colored. Non-telomeric clipped bases are transparent. Dark grey reads represent the non-telomeric end of a discordant read pair. (b) Unannotated structural variation (position 23,228,785; opaque non-telomeric clipped reads) opposite of a telomere insertion (position 23,228,744) in melanoma sample SP82836.



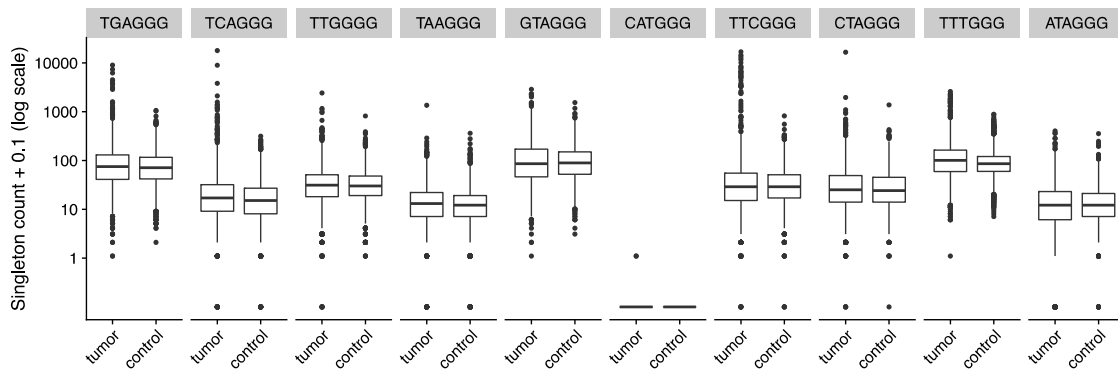
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Supplementary Figure 9: TERRA expression in tumor samples with different TMM-associated mutations. The samples are colored by the number of TERRA reads mapping to telomere insertion sites. The center line of the boxplot is the median, the bounds of the box represent the first and third quartiles, the upper and lower whiskers extend from the hinge to the largest or smallest value, respectively, no further than 1.5 * IQR from the hinge (where IQR is the inter-quartile range, or distance between the first and third quartiles). **** $p < 0.0001$, Wilcoxon rank-sum test.

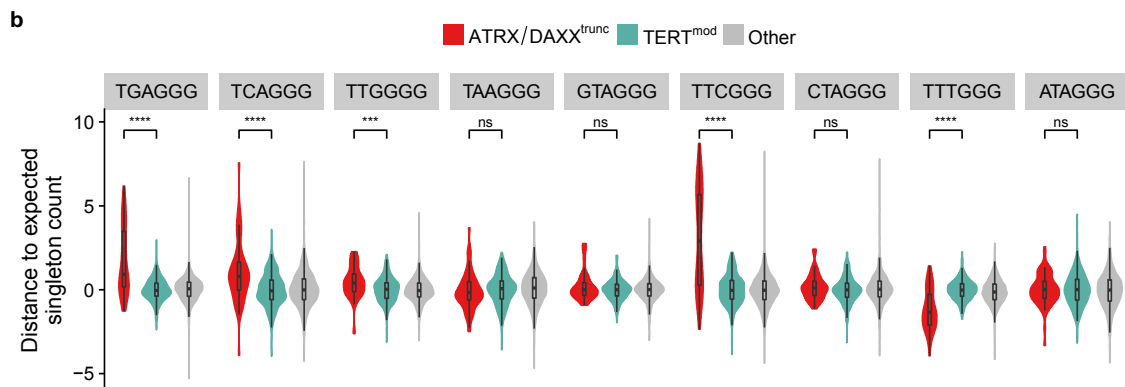
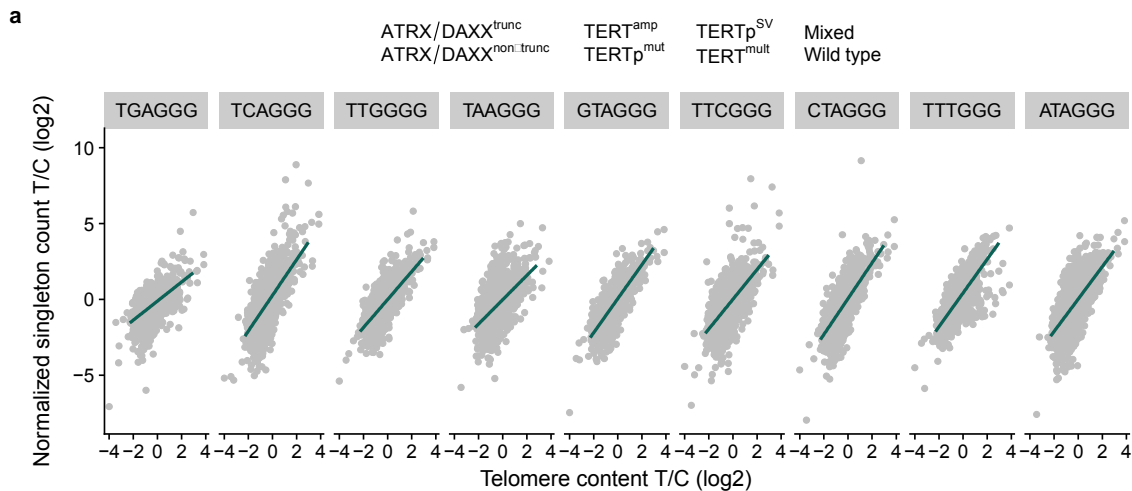




Supplementary Figure 11: The neighborhood of TVRs is indicative of the telomere maintenance mechanism. (a) Receiver operating characteristic for the classification of samples with ALT-associated mutations from telomere variant repeats. Red: no specific sequence context required. Blue: singletons ((TTAGGG)₃-NNNGGG-(TTAGGG)₃). (b) Area under the curve (AUC) for the classification of ALT using repeat type counts in different sequence context.



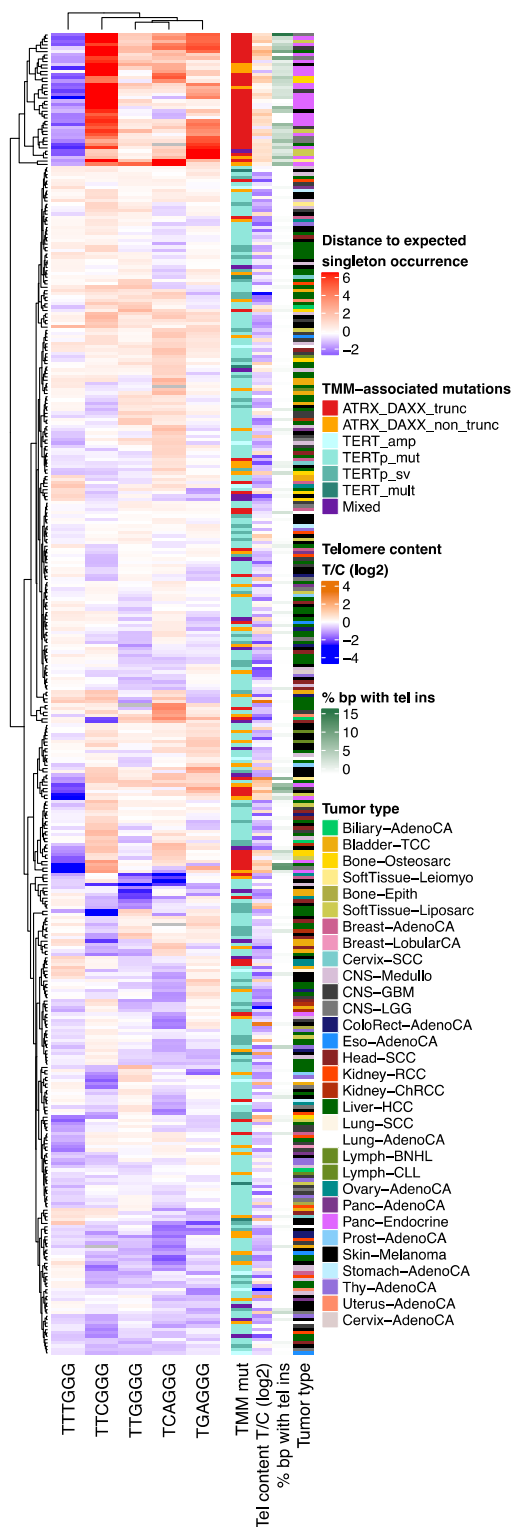
Supplementary Figure 12: Raw counts of singleton TVRs across all samples. Singleton counts are shown in log scale for all tumor and control samples. A pseudocount of 0.1 was added. The center line of the boxplot is the median, the bounds of the box represent the first and third quartiles, the upper and lower whiskers extend from the hinge to the largest or smallest value, respectively, no further than $1.5 * \text{IQR}$ from the hinge (where IQR is the inter-quartile range, or distance between the first and third quartiles).



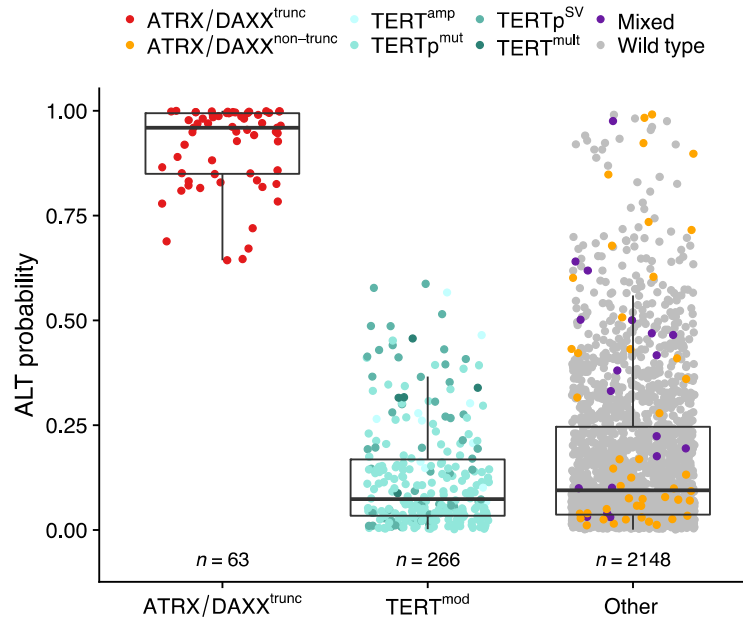
Supplementary Figure 13: Singleton TVRs. (a) Pattern count tumor/control log2 ratios of all patients plotted against telomere content tumor/control log2 ratios for analyzed singletons. The regression line through the TERT^{mod} samples is shown in green and is defined as the expected pattern count. (b) Distance to the expected singleton repeat count in $\text{ATRX/DAXX}^{\text{trunc}}$ and TERT^{mod} samples. The center line of the boxplot is the median, the bounds of the box represent the first and third quartiles, the upper and lower whiskers extend from the hinge to the largest or smallest value, respectively, no further than $1.5 * \text{IQR}$ from the hinge (where IQR is the inter-quartile range, or distance between the first and third quartiles). **** $p < 0.0001$; *** $p < 0.001$; ns = not significant, Wilcoxon rank-sum tests after Bonferroni correction.



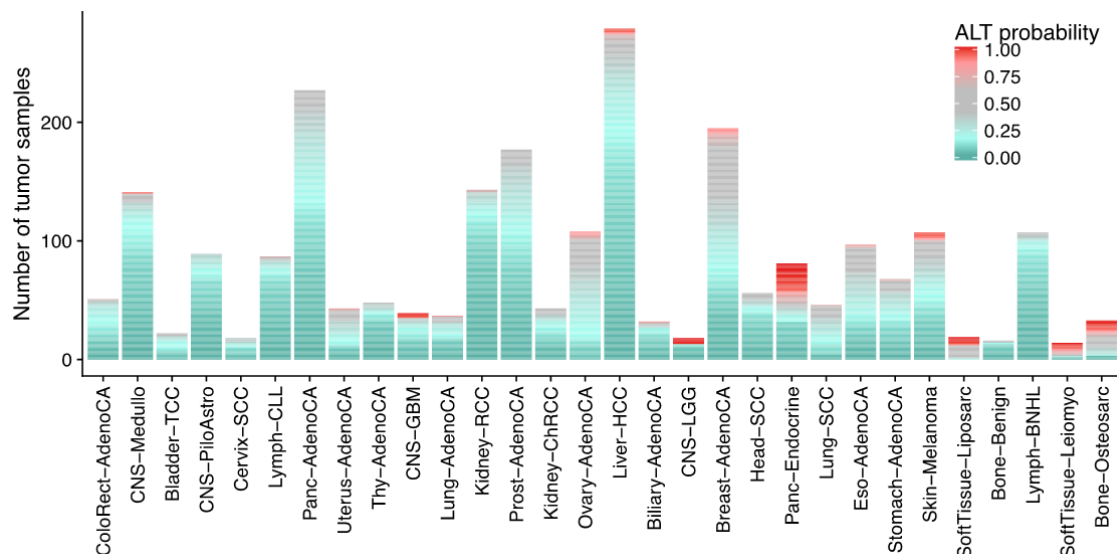
130 **Supplementary Figure 14: Correlation of singleton TVRs in ATRX/DAXX^{trunc} samples.** The
131 Spearman correlation coefficients for the occurrence of the significantly enriched/depleted singleton
132 TVRs in ATRX/DAXX^{trunc} samples is shown.
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Supplementary Figure 15: Clustering by singleton TVR occurrences. The heatmap depicts the difference of observed singleton occurrence to the expected occurrence (columns) for tumor samples with TERT^{mod} and/or mutations in ATRX or DAXX (rows). The TMM-associated mutations, telomere content tumor/control (log₂), percent of breakpoints with telomere insertion and tumor type are annotated.



Supplementary Figure 16: ALT probability of tumor samples with different TMM-associated mutations. The ALT probability was derived from a random forest classifier trained to distinguish ATRX/DAXX^{trunc} from TERT^{mod} samples based on the following features: telomere content tumor/control log2 ratio, number of telomere insertions, number of break points and the distance of TGAGGG, TCAGGG, TTGGGG, TTCGGG and TTTGGG singletons to their expected occurrence. The classifier was only applied to samples without missing data. The center line of the boxplot is the median, the bounds of the box represent the first and third quartiles, the upper and lower whiskers extend from the hinge to the largest or smallest value, respectively, no further than 1.5 * IQR from the hinge (where IQR is the inter-quartile range, or distance between the first and third quartiles).



Supplementary Figure 17: Prediction of ALT probability in different tumor types. For each tumor sample, the ALT probability predicted by a random forest classifier is shown. The tumor types are ordered by mean telomere content tumor/control log2 ratio (from left to right). Cohorts with sample sizes below 15 are not shown.

Supplementary Table 1: Mutated genes significantly associated with telomere insertions. The genes were obtained from correlation analysis of telomere insertions and mutations of genes from the TelNet database. q-values were calculated with Wilcoxon rank-sum tests with Benjamini-Hochberg correction.

Gene	q-value	Association with telomeres/telomere maintenance
<i>TP53</i>	1.9×10^{-42}	Co-mutations of the tumor suppressor gene <i>TP53</i> with <i>ATRX</i> and <i>H3F3A</i> were found in pediatric glioblastoma ¹ . p53 was also found to inhibit telomerase ² , thus truncated p53 was associated with high levels of TERT ³ .
<i>ATRX</i> / <i>DAXX</i>	2.6×10^{-6} / 0.019	The death domain-associated protein DAXX and the chromatin remodeling factor ATRX (a-thalassemia/mental retardation syndrome protein) are specifically associated with the H3.3 deposition machinery. The loss of function of ATRX and/or DAXX correlates with the ALT phenotype ⁴⁻⁷ .
<i>PLCB2</i>	7.8×10^{-4}	The yeast homologue of <i>PLCB2</i> , <i>PLC1</i> , was found in a <i>TLC1</i> knock out study ⁸ .
<i>MEN1</i>	0.017	The tumor suppressor menin, encoded by <i>MEN1</i> , was reported to negatively regulate telomerase by binding to the <i>TERT</i> promotor ⁹ , while siRNA-based inhibition of <i>MEN1</i> does not lead to the up-regulation of hTERT mRNA expression ¹⁰ . A germline mutation in <i>MEN1</i> predisposed to developing pancreatic neuroendocrine tumors. In a study of 50 patient samples, 6% of this tumor entity show loss of ATRX and/or DAXX expression and activation of ALT ⁶ . Also, it was found to inversely be associated with telomere length ¹¹ . In agreement with this, mutations in <i>MEN1</i> were associated with increased telomere length in pancreatic endocrine neoplasms ¹² .
<i>TSSC4</i>	0.017	<i>TSSC4</i> was found in close proximity to <i>RAP1</i> by a PCA/BiFC assay ¹³ .
<i>RB1</i>	0.018	The tumor suppressor <i>RB1</i> was predicted to be potentially involved in telomere maintenance by Lovejoy <i>et al.</i> because it is part of the DNA damage response machinery ⁵ . Gonzalez-Vasconcellos <i>et al.</i> showed that both telomeric chromatin compaction and telomeric repeat-containing containing RNA (TERRA) transcription are dependent on <i>RB1</i> expression ¹⁴ .
<i>ABCC8</i>	0.04	The yeast homologue of <i>ABCC8</i> , <i>YOR1</i> , was found in two independent deletions screens in <i>S. cerevisiae</i> ^{3,15} .

Supplementary Table 2: Prevalence of ALT-like telomere composition in different tumor types.
Tumor samples were considered to have an ALT-like telomere composition if the difference of the TGAGGG, TCAGGG, TTGGGG or TTCGGG singleton occurrence to the expected occurrence was larger than 2.5, or if the difference of the TTTGGG occurrence in t-type context to the expected occurrence was lower than -2.

Tumor type	n/N	%
SoftTissue-Leiomyo	9/15	60
Panc-Endocrine	34/81	42
SoftTissue-Liposarc	7/19	37
Bone-Osteosarc	10/35	29
CNS-LGG	5/18	28
CNS-GBM	5/39	13
Bone-Epith	1/10	10
Kidney-ChRCC	3/43	7
Skin-Melanoma	8/107	7
Bone-Benign	1/16	6
Liver-HCC	14/305	5
Uterus-AdenoCA	2/44	5
CNS-Medullo	6/141	4
ColoRect-AdenoCA	2/52	4
Biliary-AdenoCA	1/33	3
Kidney-RCC	4/143	3
Eso-AdenoCA	2/97	2
Lymph-BNHL	2/107	2
Lymph-CLL	2/90	2
Ovary-AdenoCA	2/110	2
Breast-AdenoCA	1/195	1
Panc-AdenoCA	2/229	1
Prost-AdenoCA	2/178	1
Bladder-TCC	0/23	0
Breast-DCIS	0/3	0
Breast-LobularCA	0/13	0
Cervix-AdenoCA	0/2	0
Cervix-SCC	0/18	0
CNS-PiloAstro	0/89	0
Head-SCC	0/56	0
Lung-AdenoCA	0/37	0
Lung-SCC	0/47	0
Myeloid-AML	0/8	0
Stomach-AdenoCA	0/68	0
Thy-AdenoCA	0/48	0

Supplementary Table 3: Feature importance in random forest classifier trained on ATRX/DAXX^{trunc} and TERT^{mod} tumor samples.

Feature	Importance
TTTGGG singleton divergence to expected count	13.59
TTCGGG singleton divergence to expected count	11.92
breakpoint count	11.01
telomere insertion count	10.03
telomere content tumor/control log2 ratio	5.34
TGAGGG singleton divergence to expected count	5.02
TCAGGG singleton divergence to expected count	3.25
TTGGGG singleton divergence to expected count	2.83

Supplementary Table 4: Histology overview. Information on the tumor types included in this study.

Abbreviation	Organ	Included subtypes
Biliary-AdenoCA	Biliary	Cholangiocarcinoma; Cholangiocarcinoma, papillary
Bladder-TCC	Bladder	Transitional cell carcinoma; Transitional cell carcinoma, papillary
Bone-Benign	Bone/SoftTissue	Osteoblastoma; Osteofibrous dysplasia; Chondroblastoma; Chondromyxoid fibroma
Bone-Epith	Bone/SoftTissue	Adamantinoma; Chordoma
Bone-Osteosarc	Bone/SoftTissue	Osteosarcoma
Breast-AdenoCA	Breast	Infiltrating duct carcinoma; Intraductal papillary adenocarcinoma with invasion; Medullary carcinoma; Mucinous adenocarcinoma
Breast-DCIS	Breast	Duct micropapillary carcinoma
Breast-LobularCA	Breast	Lobular carcinoma
Cervix-AdenoCA	Cervix	Adenocarcinoma
Cervix-SCC	Cervix	Squamous Cell Carcinoma
CNS-GBM	CNS	Glioblastoma
CNS-LGG	CNS	Oligodendroglioma; ATRX-mutant, 1p/19q-intact lower grade gliomas
CNS-Medullo	CNS	Desmoplastic medulloblastoma; Large cell medulloblastoma; Medulloblastoma
CNS-PiloAstro	CNS	Pilocytic astrocytoma
ColoRect-AdenoCA	Colon/Rectum	Adenocarcinoma; Adenocarcinoma, mucinous; Mucinous adenocarcinoma
Eso-AdenoCA	Esophagus	Adenocarcinoma
Head-SCC	Head/Neck	Squamous cell carcinoma
Kidney-ChRCC	Kidney	Adenocarcinoma, chromophobe type
Kidney-RCC	Kidney	Adenocarcinoma, clear cell type; Adenocarcinoma, papillary type
Liver-HCC	Liver	Combined hepatocellular + cholangiocarcinoma; Fibrolamellar hepatocellular carcinoma; Hepatocellular carcinoma
Lung-AdenoCA	Lung	Adenocarcinoma, in situ; Adenocarcinoma, invasive; Adenocarcinoma, invasive, mucinous
Lung-SCC	Lung	Basaloid squamous cell carcinoma; Squamous cell carcinoma
Lymph-BNHL	Lymphoid	Burkitt lymphoma; Diffuse large B-cell lymphoma; Follicular lymphoma; Marginal zone B-cell lymphoma; Post-transplant lymphoproliferative disorder, early lesion
Lymph-CLL	Lymphoid	Chronic lymphocytic leukemia
Myeloid-AML	Myeloid	Acute myeloid leukemia
Ovary-AdenoCA	Ovary	Adenocarcinoma; Serous cystadenocarcinoma
Panc-AdenoCA	Pancreas	Acinar cell carcinoma; Adenocarcinoma; Adenocarcinoma, mucinous; Carcinoma, adenosquamous; Invasive carcinoma arising in IPMN; Pancreatic ductal carcinoma
Panc-Endocrine	Pancreas	Neuroendocrine carcinoma
Prost-AdenoCA	Prostate	Adenocarcinoma
Skin-Melanoma	Skin	Malignant melanoma
SoftTissue-Leiomyo	Bone/SoftTissue	Leiomyosarcoma
SoftTissue-Liposarc	Bone/SoftTissue	Liposarcoma
Stomach-AdenoCA	Stomach	Adenocarcinoma; Adenocarcinoma, mucinous; Adenocarcinoma, papillary; Adenocarcinoma, poorly cohesive; Adenocarcinoma, tubular
Thy-AdenoCA	Thyroid	Adenocarcinoma, classical type; Adenocarcinoma, columnar cell type; Adenocarcinoma, follicular type
Uterus-AdenoCA	Uterus	Adenocarcinoma, endometrioid; Serous cystadenocarcinoma

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Supplementary Information:

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Novel somatic mutation calling methods

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Drivers and functional interpretation

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Integration of transcriptome and genome

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Integration of epigenome and genome

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Patterns of structural variations, signatures, genomic correlations, retrotransposons, mobile elements

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Mutation signatures and processes

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Germline cancer genome

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Tumour subtypes and clinical translation

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Evolution and heterogeneity

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Cmero^{374,375,376}, Yupeng Cun³⁷⁷, Kevin J Dawson², Jonas Demeulemeester^{63,64}, Stefan C Dentro^{2,64,354}, Amit G Deshwar³⁷⁸, Nilgun Donmez^{151,159}, Ruben M Drews²⁹⁴, Roland Eils^{52,54,66,67}, Yu Fan¹⁴⁸, Matthew W Fittall⁶⁴, Dale W Garsed^{187,188}, Moritz Gerstung^{7,8}, Gad Getz^{3,4,5,6}, Santiago Gonzalez^{7,8}, Gavin Ha³, Kerstin Haase⁶⁴, Marcin Imielinski^{299,300}, Lara Jerman^{8,379}, Yuan Ji^{380,381}, Clemency Jolly⁶⁴, Kortine Kleinheinz^{52,54}, Juhee Lee³⁸², Henry Lee-Six², Ignaty Leshchiner³, Dimitri Livitz³, Geoff Macintyre²⁹⁴, Salem Malikic^{151,159}, Florian Markowetz^{294,295}, Iñigo Martincorena², Thomas J Mitchell^{2,295,383}, Quaid D Morris^{358,384}, Ville Mustonen^{320,321,322}, Layla Oesper³⁸⁵, Martin Peifer³⁷⁷, Myron Peto³⁸⁶, Benjamin J Raphael¹²⁰, Daniel Rosebrock³, Yulia Rubanova^{160,358}, S Cenk Sahinalp^{151,158,159}, Adriana Salcedo⁹, Matthias Schlesner^{52,110}, Steven E Schumacher^{3,217}, Subhajt Sengupta³⁸⁷, Ruian Shi³⁸⁴, Seung Jun Shin²⁶⁴, **Paul T Spellman**³⁸⁸, Oliver Spiro³, Lincoln D Stein^{9,10}, Maxime Tarabichi^{2,64}, **Peter Van Loo**^{63,64}, Shankar Vembu^{384,389}, Ignacio Vázquez-García^{2,167,327,328}, Wenyi Wang¹⁴⁸, **David C Wedge**^{2,354,355}, David A Wheeler^{164,165}, Jeffrey A Wintersinger^{192,358,390}, Tsun-Po Yang³⁷⁷, Xiaotong Yao^{299,316}, Kaixian Yu³⁹¹, Ke Yuan^{294,369,372} and Hongtu Zhu^{392,393}

Portals, visualisation and software infrastructure

Fatima Al-Shahrour³⁵⁷, Elisabet Barrera⁷, Wojciech Bazant⁷, Alvis Brazma⁷, Isidro Cortés-Ciriano^{237,238,239}, Brian Craft²⁴⁰, David Craft³, Vincent Ferretti^{45,69}, Nuno A Fonseca^{7,70}, Anja Füllgrabe⁷, Mary J Goldman²⁴⁰, **David Haussler**^{240,394}, Wolfgang Huber⁸, Maria Keays⁷, Alfonso Muñoz⁷, Brian D O'Connor^{45,50}, Irene Papatheodorou⁷, Robert Petryszak⁷, Elena Piñeiro-Yáñez³⁵⁷, Alfonso Valencia^{105,111}, **Miguel Vazquez**^{105,112}, John N Weinstein^{395,396}, Qian Xiang¹¹⁶, Junjun Zhang⁴⁵ and **Jingchun Zhu**²⁴⁰

Mitochondrial variants and HLA/immunogenicity

Peter J Campbell^{1,2}, Yiwen Chen¹⁴⁸, Chad J Creighton²⁴¹, Li Ding^{138,139,146}, Akihiro Fujimoto⁴⁸, Masashi Fujita⁴⁸, Gad Getz^{3,4,5,6}, Leng Han²³¹, Takanori Hasegawa⁸⁷, Shuto Hayashi⁸⁷, Seiya Imoto^{86,87}, Young Seok Ju^{2,181}, Hyung-Lae Kim²⁷, Youngwook Kim^{96,97}, Youngil Koh^{307,308}, Mitsuhiro Komura⁸⁷, Jun Li¹⁴⁸, **Han Liang**³⁹⁷, Iñigo Martincorena², Satoru Miyano⁸⁷, Shinichi Mizuno³⁹⁸, **Hidewaki Nakagawa**⁴⁸, Keunchil Park^{206,207}, Eigo Shimizu⁸⁷, Yumeng Wang^{148,399}, John N Weinstein^{395,396}, Yanxun Xu⁴⁰⁰, Rui Yamaguchi⁸⁷, Fan Yang³⁸⁴, Yang Yang²³¹, Christopher J Yoon¹⁸¹, Sung-Soo Yoon³⁰⁸, Yuan Yuan¹⁴⁸, Fan Zhang²⁴⁶ and Zemin Zhang^{246,271}

Pathogens

Malik Alawi^{401,402}, Ivan Borozan⁹, Daniel S Brewer^{403,404}, Colin S Cooper^{404,405,406}, Nikita Desai⁴⁵, **Roland Eils**^{52,54,66,67}, Vincent Ferretti^{45,69}, Adam Grundhoff^{401,407}, Murat Iskar⁴⁰⁸, Kortine Kleinheinz^{52,54}, Peter Lichter⁴⁰⁸, **Hidewaki Nakagawa**⁴⁸, Akinyemi I Ojesina^{255,256,257}, Chandra Sekhar Pedomallu^{3,6,172}, Matthias Schlesner^{52,110}, Xiaoping Su¹⁴⁴ and Marc Zapatka⁴⁰⁸

Tumour Specific Providers – Australia (Ovarian cancer)

Kathryn Alsop^{409,410}, Australian Ovarian Cancer Study Group^{187,311,411}, **David D L Bowtell**^{187,291}, Timothy JC Bruxner¹⁸⁵, Angelika N Christ¹⁸⁵, Elizabeth L Christie¹⁸⁷, Stephen M Cordner⁴¹², Prue A Cowin¹⁸⁷, Ronny Drapkin⁴¹³, Dariush Etemadmoghadam^{187,188}, Sian Fereday⁴¹⁴, Dale W Garsed^{187,188}, Joshy George¹⁷⁰, Sean M Grimmond³⁶³, Anne Hamilton¹⁸⁷, Oliver Holmes^{311,312}, Jillian A Hung^{415,416}, Karin S Kassahn^{185,417}, Stephen H Kazakoff^{311,312}, Catherine J Kennedy^{418,419}, Conrad R Leonard^{311,312}, Linda Miles¹⁸⁷, David K Miller^{185,360,420}, Gisela Mir Arnau¹⁸⁷, Chris Mitchell¹⁸⁷, Felicity Newell^{311,312}, Katia Nones^{311,312}, Ann-Marie Patch^{311,312}, John V Pearson^{311,312}, Michael C Quinn^{311,312}, Mark Shackleton^{188,218}, Darrin F Taylor¹⁸⁵, Heather Thorne¹⁸⁷, Nadia Traficante¹⁸⁷, Ravikiran Vedururu¹⁸⁷, Nick M Waddell³¹², Nicola Waddell^{311,312}, Paul M Waring²⁵³, Scott Wood^{311,312}, Qinying Xu^{311,312} and Anna deFazio^{421,422,423}

Tumour Specific Providers – Australia (Pancreatic cancer)

Matthew J Anderson¹⁸⁵, Davide Antonello⁴²⁴, Andrew P Barbour^{425,426}, Claudio Bassi⁴²⁴, Samantha Bersani⁴²⁷, **Andrew V Biankin**^{359,360,361,362}, Timothy JC Bruxner¹⁸⁵, Ivana Cataldo^{427,428}, David K Chang^{360,362}, Lorraine A Chantrill³⁶⁰, Yoke-Eng Chiew⁴²¹, Angela Chou^{360,429}, Angelika N Christ¹⁸⁵, Sara Cingarlini³⁷, Nicole Cloonan⁴³⁰, Vincenzo Corbo^{428,431, 432}, Fraser R Duthie^{433,434}, J Lynn Fink^{105,185}, Anthony J Gill^{360,435}, Janet S Graham^{362,436}, **Sean M Grimmond**³⁶³, Ivon Harliwong¹⁸⁵, Oliver Holmes^{311,312}, Nigel B Jamieson^{361,362,437}, Amber L Johns^{360,420}, Karin S Kassahn^{185,417}, Stephen H Kazakoff^{311,312}, James G Kench^{360,435,438}, Luca Landoni⁴²⁴, Rita T Lawlor⁴²⁸, Conrad R Leonard^{311,312}, Andrea Mafficini⁴²⁸, Neil D Merrett^{424,439}, David K Miller^{185,360,420}, Marco Miotto⁴²⁴, Elizabeth A Musgrove³⁶², Adnan M Nagrial³⁶⁰, Felicity Newell^{311,312}, Katia Nones^{311,312}, Karin A Oien^{253,440}, Marina Pajic³⁶⁰, Ann-Marie Patch^{311,312}, John V Pearson^{311,312}, Mark Pinese³⁶⁰, Andreia V Pinho³⁶⁰, Michael C Quinn^{311,312}, Alan J Robertson¹⁸⁵, Ilse Rooman³⁶⁰, Borislav C Rusev⁴²⁸, Jaswinder S Samra^{424,435}, Maria Scardoni⁴²⁷, Christopher J Scarlett^{360,441}, Aldo Scarpa⁴²⁸, Elisabetta Sereni⁴²⁴, Katarzyna O Sikora⁴²⁸, Michele Simbolo⁴³¹, Morgan L Taschuk⁴⁵, Christopher W Toon³⁶⁰, Giampaolo Tortora^{37,38}, Caterina Vicentini⁴²⁸, Nick M Waddell³¹², Nicola Waddell^{311,312}, Scott Wood^{311,312}, Jianmin Wu³⁶⁰, Qinying Xu^{311,312} and Nikolajs Zeps⁴⁴²

Tumour Specific Providers – Australia (Skin cancer)

Lauri A Aaltonen⁴⁴³, Andreas Behren⁴⁴⁴, Hazel Burke⁴⁴⁵, Jonathan Cebon⁴⁴⁴, Rebecca A Dagg⁴⁴⁶, Ricardo De Paoli-Iseppi⁴⁴⁷, Ken Dutton-Regester³¹¹, Matthew A Field⁴⁴⁸, Anna Fitzgerald⁴⁴⁹, Sean M Grimmond³⁶³, **Nicholas K Hayward**^{311,445}, Peter Hersey⁴⁴⁵, Oliver Holmes^{311,312}, Valerie Jakrot⁴⁴⁵, Peter A Johansson³¹¹, Hojabr Kakavand⁴⁴⁷, Stephen H Kazakoff^{311,312}, Richard F Kefford⁴⁵⁰, Loretta MS Lau⁴⁵¹, Conrad R Leonard^{311,312}, Georgina V Long⁴⁵², **Graham J Mann**^{453,454}, Felicity Newell^{311,312}, Katia Nones^{311,312}, Ann-Marie Patch^{311,312}, John V Pearson^{311,312}, Hilda A Pickett⁴⁵¹, Antonia L Pritchard³¹¹, Gulietta M Pupo⁴⁵⁵, Robyn PM Saw⁴⁵²,

Sarah-Jane Schramm⁴⁵⁶, **Richard A Scolyer**^{#422,452,457,458}, Mark Shackleton^{188,218}, Catherine A Shang⁴⁵⁹, Ping Shang⁴⁵², Andrew J Spillane⁴⁵², Jonathan R Stretch⁴⁵², Varsha Tembe⁴⁵⁶, John F Thompson⁴⁵², Ricardo E Vilain⁴⁵⁷, Nick M Waddell³¹², Nicola Waddell^{311,312}, James S Wilmott⁴⁵², Scott Wood^{311,312}, Qinying Xu^{311,312} and Jean Y Yang⁴⁶⁰

Tumour Specific Providers – Canada (Pancreatic cancer)

John Bartlett^{461,462}, Prashant Bavi⁴⁶³, Ivan Borozan⁹, Dianne E Chadwick⁴⁶⁴, Michelle Chan-Seng-Yue⁴⁶³, Sean Cleary^{463,465}, Ashton A Connor^{466,467}, Karolina Czajka⁴⁶⁸, Robert E Denroche⁴⁶³, Neesha C Dhani⁴⁶⁹, Jenna Eagles⁷⁹, Vincent Ferretti^{45,69}, Steven Gallinger^{463,466,467}, Robert C Grant^{463,470}, David Hedley⁴⁶⁹, Michael A Hollingsworth⁴⁷¹, **Thomas J Hudson**^{#78,79}, Gun Ho Jang⁴⁶³, Jeremy Johns⁷⁹, Sangeetha Kalimuthu⁴⁶³, Sheng-Ben Liang⁴⁷², Ilinca Lungu^{463,473}, Xuemei Luo⁹, Faridah Mbabaali⁷⁹, **John D McPherson**^{#79,463,474}, Treasa A McPherson⁴⁷⁰, Jessica K Miller⁷⁹, Malcolm J Moore⁴⁶⁹, Faiyaz Notta^{463,475}, Danielle Pasternack⁷⁹, Gloria M Petersen⁴⁷⁶, Michael H A Roehrl^{133,463,477,478,479}, Michelle Sam⁷⁹, Iris Selander⁴⁷⁰, Stefano Serra²⁵³, Sagedeh Shahabi⁴⁷², **Lincoln D Stein**^{#9,10}, Morgan L Taschuk⁴⁵, Sarah P Thayer¹⁰⁶, Lee E Timms⁷⁹, Gavin W Wilson^{9,463}, Julie M Wilson⁴⁶³ and Bradly G Wouters⁴⁸⁰

Tumour Specific Providers – Canada (Prostate cancer)

Timothy A Beck⁴⁵, Vinayak Bhandari⁹, Paul C Boutros^{9,133,142,143}, **Robert G Bristow**^{#133,481,482,483,484}, Colin C Collins¹⁵¹, Shadrielle MG Espiritu⁹, Neil E Fleshner⁴⁸⁵, Natalie S Fox⁹, Michael Fraser⁹, Syed Haider⁹, Lawrence E Heisler⁴⁸⁶, Vincent Huang⁹, Emilie Lalonde⁹, Julie Livingstone⁹, John D McPherson^{79,463,474}, Alice Meng⁴⁸⁷, Veronica Y Sabelnykova⁹, Adriana Salcedo⁹, Yu-Jia Shiah⁹, Theodorus Van der Kwast⁴⁸⁸ and Takafumi N Yamaguchi⁹

Tumour Specific Providers – China (Gastric cancer)

Shuai Ding⁴⁸⁹, Daiming Fan⁴⁹⁰, Yong Hou^{39,249}, Yi Huang^{153,154}, Lin Li³⁹, Siliang Li^{39,249}, Dongbing Liu^{39,249}, Xingmin Liu^{39,249}, **Youyong Lu**^{#28,29,30}, Yongzhan Nie^{490,491}, Hong Su^{39,249}, Jian Wang³⁹, Kui Wu^{39,249}, Xiao Xiao¹⁵⁴, Rui Xing^{29,492}, **Huanming Yang**^{#39}, Shanlin Yang⁴⁸⁹, Yingyan Yu^{493, 230}, Xiuqing Zhang³⁹, Yong Zhou³⁹ and Shida Zhu^{39,249}

Tumour Specific Providers – EU: France (Renal cancer)

Rosamonde E Banks⁴⁹⁴, Guillaume Bourque^{495,496}, Alvis Brazma⁷, Paul Brennan⁴⁹⁷, **Mark Lathrop**^{#496}, Louis Letourneau⁴⁹⁸, Yasser Riazalhosseini⁴⁹⁶, Ghislaine Scelo⁴⁹⁷, **Jörg Tost**^{#499}, Naveen Vasudev⁵⁰⁰ and Juris Viksna⁵⁰¹

Tumour Specific Providers – EU: United Kingdom (Breast cancer)

Sung-Min Ahn⁵⁰², Ludmil B Alexandrov^{2,317}, Samuel Aparicio⁵⁰³, Laurent Arnould⁵⁰⁴, MR Aure⁵⁰⁵, Shriram G Bhosle², E Birney⁷, Ake Borg⁵⁰⁶, S Boyault⁵⁰⁷, AB Brinkman⁵⁰⁸, JE Brock⁵⁰⁹, A Broeks⁵¹⁰, Adam P Butler², AL Børresen-Dale⁵⁰⁵, C Caldas^{511,512}, Peter J Campbell^{1,2}, Suet-Feung Chin^{511,512}, Helen Davies², C Desmedt⁵¹³, L Dirix⁵¹⁴, S Dronov², Anna Ehinger⁵¹⁵, JE Eyfjord⁵¹⁶, GG Van den Eynden⁵¹⁷, A Fatima²¹⁷, Jorge Reis Filho⁵¹⁸, JA Foekens⁵¹⁹, PA Futreal⁵²⁰, Øystein Garred^{521,522}, Moritz Gerstung^{7,8}, Dilip D Giri⁵¹⁸, D Glodzik², Dorte Grabau⁵²³, Holmfridur Hilmarisdottir⁵¹⁶, GK Hooijer⁵²⁴, Jocelyne Jacquemier⁵²⁵, SJ Jang⁵²⁶, Jon G Jonasson⁵¹⁶, Jos Jonkers⁵²⁷, HY Kim⁵²⁵, Tari A King^{528,529}, Stian Knappskog², G Kong⁵²⁵, S Krishnamurthy⁵³⁰, S Van Laere⁵¹⁴, SR Lakhani⁵³¹, A Langerød⁵⁰⁵, Denis Larsimont⁵³², HJ Lee⁵²⁶, JY Lee⁵³³, Ming Ta Michael Lee⁵²⁰, Yilong Li², Ole Christian Lingjærde⁵³⁴, Gaetan MacGrogan⁵³⁵, JW Martens⁵³⁶, Sancha Martin^{2,369}, Iñigo Martincorena², Andrew Menzies², Sandro Morganella², Ville Mustonen^{320,321,322}, Serena Nik-Zainal^{2,324,325,326}, Sarah O'Meara², I Pauporté¹⁸, Sarah Pinder⁵³⁷, X Pivot⁵³⁸, Elena Provenzano⁵³⁹, CA Purdie⁵⁴⁰, Keiran M Raine², M Ramakrishna², K Ramakrishnan², AL Richardson²¹⁷, M Ringnér⁵⁰⁶, Javier Bartolomé Rodríguez¹⁰⁵, FG Rodríguez-González¹⁷⁵, G Romieu⁵⁴¹, Roberto Salgado²⁵³, Torill Sauer⁵³⁴, R Shepherd², AM Sieuwerts¹⁷⁷, PT Simpson⁵³¹, M Smid⁵⁴², C Sotiriou⁵⁵, PN Span⁵⁴³, J Staaf⁵⁰⁶, Lucy Stebbings², Ólafur Andri Stefánsson⁵⁴⁴, Alasdair Stenhouse⁵⁴⁵, **Michael Rudolf Stratton**^{#2}, HG Stunnenberg^{249,546}, Fred Sweep⁵⁴⁷, BK Tan⁵⁴⁸, Jon W Teague², Gilles Thomas⁵⁴⁹, AM Thompson⁵⁴⁵, S Tommasi⁵⁵⁰, I Treilleux^{551,552}, Andrew Tutt²¹⁷, NT Ueno³⁹³, Peter Van Loo^{63,64}, P Vermeulen⁵¹⁴, Alain Viari⁴²⁸, MJ van de Vijver²⁵³, A Vincent-Salomon⁵⁴⁶, David C Wedge^{2,354,355}, Bernice Huimin Wong⁵⁵³, Lucy Yates², X Zou², CHM van Deurzen⁵³⁶ and L van't Veer^{554,555}

Tumour Specific Providers – Germany (Malignant lymphoma)

Ole Ammerpohl^{556,557}, Sietse Aukema^{558,559}, Anke K Bergmann⁵⁶⁰, Stephan H Bernhart^{276,277,281}, Hans Binder^{276,277}, Arndt Borkhardt⁵⁶¹, Christoph Borst⁵⁶², Benedikt Brors^{82,119,278}, Birgit Burkhardt⁵⁶³, Alexander Claviez⁵⁶⁴, Roland Eils^{52,54,66,67}, Maria Elisabeth Goebler⁵⁶⁵, Andrea Haake⁵⁵⁶, Siegfried Haas⁵⁶², Martin Hansmann⁵⁶⁶, Jessica I Hoell⁵⁶¹, Steve Hoffmann^{277,279,280,281}, Michael Hummel⁵⁶⁷, Daniel Hübschmann^{54,66,83,84,85}, Dennis Karsch⁵⁶⁸, Wolfram Klapper⁵⁵⁹, Kortine Kleinheinz^{52,54}, Michael Kneba⁵⁶⁸, Jan O Korbel^{7,8}, Helene Kretzmer^{277,281}, Markus Kreuz⁵⁶⁹, Dieter Kube⁵⁷⁰, Ralf Küppers⁵⁷¹, Chris Lawerenz⁶⁷, Dido Lenze⁵⁶⁷, Peter Lichter⁴⁰⁸, Markus Loeffler⁵⁶⁹, Cristina López^{262,556}, Luisa Mantovani-Löffler⁵⁷², Peter Möller⁵⁷³, German Ott⁵⁷⁴, Bernhard Radlwimmer⁴⁰⁸, Julia Richter^{556,559}, Marius Rohde⁵⁷⁵, Philip C Rosenstiel⁵⁷⁶, Andreas Rosenwald⁵⁷⁷, Markus B Schilhabel⁵⁷⁶, Matthias Schlesner^{52,110}, Stefan Schreiber⁵⁷⁸, **Reiner Siebert**^{#261,262}, Peter F Stadler^{276,277,281}, Peter Staib⁵⁷⁹, Stephan Stilgenbauer⁵⁸⁰, Stephanie Sungalee⁸, Monika Szczepanowski⁵⁵⁹, Umut H Toprak^{54,581}, Lorenz HP Trümper⁵⁷⁰, Rabea Wagener^{262,556} and Thorsten Zenz⁸²

Tumour Specific Providers – Germany (Pediatric Brain cancer)

Ivo Buchhalter^{52,53,54}, Juergen Eils^{66,67}, Roland Eils^{52,54,66,67}, Volker Hovestadt⁴⁰⁸, Barbara Hutter^{80,81,82}, David TW Jones^{301,302}, Natalie Jäger⁵², Christof von Kalle⁸⁴, Marcel Kool^{98,301}, Jan O Korbel^{7,8}, Andrey Korshunov⁹⁸, Pablo Landgraf⁵⁸², Chris Lawrenz⁶⁷, Hans Lehrach⁵⁸³, **Peter Lichter**^{#408}, Paul A Northcott⁵⁸⁴, Stefan M Pfister^{98,301,585}, Bernhard Radlwimmer⁴⁰⁸, Guido Reifenberger⁵⁸², Matthias Schlesner^{52,110}, Hans-Jörg Warnatz⁵⁸³, Joachim Weischenfeldt^{8,113,114}, Stephan Wolf⁵⁸⁶, Marie-Laure Yaspo⁵⁸³ and Marc Zapatka⁴⁰⁸

Tumour Specific Providers – Germany (Prostate cancer)

Yassen Assenov⁵⁸⁷, Benedikt Brors^{82,119,278}, Juergen Eils^{66,67}, Roland Eils^{52,54,66,67}, Lars Feuerbach¹¹⁹, Clarissa Gerhauser²⁸⁵, Jan O Korbel^{7,8}, Chris Lawrenz⁶⁷, Hans Lehrach⁵⁸³, Sarah Minner⁵⁸⁸, Christoph Plass²⁸⁵, **Guido Sauter**^{#589}, Thorsten Schlomm^{114,590}, Nikos Sidiropoulos¹¹³, Ronald Simon⁵⁸⁹, **Holger Sültmann**^{#82,591}, Hans-Jörg Warnatz⁵⁸³, Dieter Weichenhan²⁸⁵, Joachim Weischenfeldt^{8,113,114} and Marie-Laure Yaspo⁵⁸³

Tumour Specific Providers – India (Oral cancer)

Nidhan K Biswas⁵⁹², Luca Landoni⁴²⁴, Arindam Maitra⁵⁹², **Partha P Majumder**^{#592} and **Rajiv Sarin**^{#593}

Tumour Specific Providers – Italy (Pancreatic cancer)

Davide Antonello⁴²⁴, Stefano Barbi⁴³¹, Claudio Bassi⁴²⁴, Samantha Bersani⁴²⁷, Giada Bonizzato⁴²⁸, Cinzia Cantù⁴²⁸, Ivana Cataldo^{427,428}, Sara Cingarlini³⁷, Vincenzo Corbo^{428,431, 432}, Angelo P Dei Tos⁵⁹⁴, Matteo Fassan⁵⁹⁵, Sonia Grimaldi⁴²⁸, Luca Landoni⁴²⁴, Rita T Lawlor⁴²⁸, Claudio Luchini⁴²⁷, Andrea Mafficini⁴²⁸, Giuseppe Malleo⁴²⁴, Giovanni Marchegiani⁴²⁴, Michele Milella³⁷, Marco Miotto⁴²⁴, Salvatore Paiella⁴²⁴, Antonio Pea⁴²⁴, Paolo Pederzoli⁴²⁴, Borislav C Rusev⁴²⁸, Andrea Ruzzenente⁴²⁴, Roberto Salvia⁴²⁴, Maria Scardoni⁴²⁷, **Aldo Scarpa**^{#428}, Elisabetta Sereni⁴²⁴, Michele Simbolo⁴³¹, Nicola Sperandio⁴²⁸, Giampaolo Tortora^{37,38} and Caterina Vicentini⁴²⁸

Tumour Specific Providers – Japan (Biliary tract cancer)

Yasuhito Arai³³, Natsuko Hama³³, Nobuyoshi Hiraoka⁵⁹⁶, Fumie Hosoda^{33,597}, Mamoru Kato³⁶⁶, Hiromi Nakamura³³, Hidenori Ojima⁵⁹⁸, Takuji Okusaka⁵⁹⁹, **Tatsuhiro Shibata**^{#33,34}, Yasushi Totoki³³ and Tomoko Urushidate³⁴

Tumour Specific Providers – Japan (Gastric cancer)

Hiroyuki Aburatani#²⁷², Yasuhito Arai³³, Masashi Fukayama⁶⁰⁰, Natsuko Hama³³, Fumie Hosoda^{33,597}, Shumpei Ishikawa⁶⁰¹, Hitoshi Katai⁶⁰², Mamoru Kato³⁶⁶, Hiroto Katoh⁶⁰³, Daisuke Komura⁶⁰¹, Genta Nagae^{272,284}, Hiromi Nakamura³³, Hirofumi Rokutan⁶⁰⁴, Mihoko Saito-Adachi³³, **Tatsuhiko Shibata#**^{33,34}, Akihiro Suzuki^{272,605}, Hirokazu Taniguchi⁶⁰⁶, Kenji Tatsuno²⁷², Yasushi Totoki³³, Tetsuo Ushiku⁶⁰⁰, Shinichi Yachida^{33,607} and Shogo Yamamoto²⁷²

Tumour Specific Providers – Japan (Liver cancer)

Hiroyuki Aburatani²⁷², Hiroshi Aikata⁶⁰⁸, Koji Arihiro⁶⁰⁸, Shun-ichi Ariizumi⁶⁰⁹, Keith A Boroevich^{47,48}, Kazuaki Chayama⁶⁰⁸, Akihiro Fujimoto⁴⁸, Masashi Fujita⁴⁸, Mayuko Furuta⁴⁸, Kunihito Gotoh⁶¹⁰, Natsuko Hama³³, Takanori Hasegawa⁸⁷, Shinya Hayami⁶¹¹, Shuto Hayashi⁸⁷, Satoshi Hirano⁶¹², Seiya Imoto^{86,87}, Mamoru Kato³⁶⁶, Yoshiiku Kawakami⁶⁰⁸, Kazuhiro Maejima⁴⁸, Satoru Miyano⁸⁷, Genta Nagae^{272,284}, **Hidewaki Nakagawa#**⁴⁸, Hiromi Nakamura³³, Toru Nakamura⁶¹², Kaoru Nakano⁴⁸, Hideki Ohdan⁶⁰⁸, Aya Sasaki-Oku⁴⁸, **Tatsuhiko Shibata#**^{33,34}, Yuichi Shiraishi⁸⁷, Hiroko Tanaka⁸⁷, Yasushi Totoki³³, Tatsuhiko Tsunoda^{47,220,221,222}, Masaki Ueno⁶¹¹, Rui Yamaguchi⁸⁷, Masakazu Yamamoto⁶⁰⁹ and Hiroki Yamaue⁶¹¹

Tumour Specific Providers – Singapore (Biliary tract cancer)

Su Pin Choo⁶¹³, Ioana Cutcutache^{267,319}, Narong Khuntikeo^{424,614}, John R McPherson^{267,319}, Choon Kiat Ong⁶¹⁵, Chawalit Pairojkul²⁵³, Irinel Popescu⁶¹⁶, **Steven G Rozen#**^{267,268,319}, **Patrick Tan#**^{254,266,267,268} and **Bin Tean Teh#**^{266,267,268,269,270}

Tumour Specific Providers – South Korea (Blood cancer)

Keun Soo Ahn⁶¹⁷, Hyung-Lae Kim²⁷, Youngil Koh^{307,308} and **Sung-Soo Yoon#**³⁰⁸

Tumour Specific Providers – Spain (Chronic Lymphocytic Leukemia)

Marta Aymerich⁶¹⁸, **Elias Campo#**^{619,620}, Josep Ll Gelpi^{46,71}, Ivo G Gut^{135,136}, Marta Gut^{135,136}, Armando Lopez-Guillermo⁶²¹, Carlos López-Otín⁶²², Xose S Puente⁶²³, Romina Royo¹⁰⁵ and David Torrents^{105,111}

Tumour Specific Providers – United Kingdom (Bone cancer)

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Tumour Specific Providers – United Kingdom (Chronic myeloid disorders)

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